

Statistical Analysis of Subcellular Proteins in Microscopy Imagery

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Joint Work with
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Motivating Applications: Radiation Biology

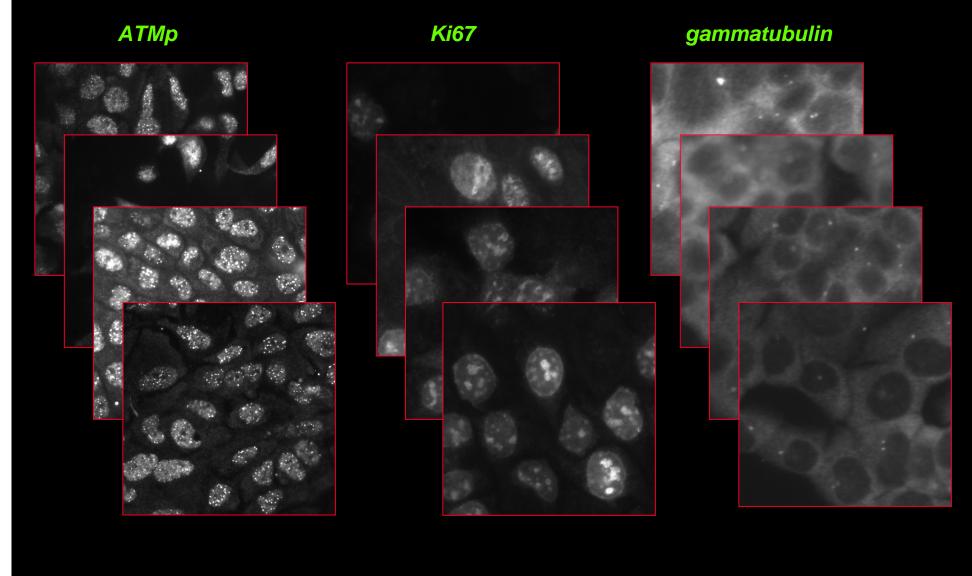


Problem: Quantify effects of radiation on cells by analyzing presence of proteins known to play crucial roles in DNA repair.

Data: Proteins in cell cultures are tagged with fluorescent probes, stained, irradiated, imaged.

Biological Challenge: High Throughput Microscopy Analysis





Sources of Image Variation



- Radiation dose/quality: high vs. low, Xray photons (sparse) vs. heavy charged particles (dense)
- Microscopy: fluorescence, confocal, deconvolution
- Time course
- Cell type and stage of mitotic cycle
- Direct vs. indirect effects of radiation
- Sample preparation

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early foci



late foci

(Petrini & Stracker, 2003)

Mathematical Challenge



- Current microscopy analysis: easy to design ad hoc, heuristic methods to get quick scientific results.
- Disadvantage: special purpose tools are designed for each new biological research question.

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- Disadvantage: special purpose tools are designed for each new biological research question.
- Mathematical models: can yield general solutions to a greater range of scientific questions and data.
- Future microscopy analysis: must move away from patchwork of image processing tools and toward principled statistical modeling and analysis.

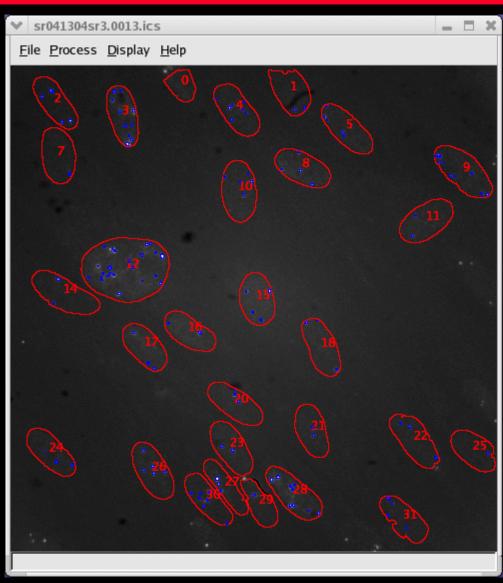
Outline



- 1. Microscopy Analysis: Biological and Mathematical Challenges
- 2. Current Approach: Microscopy Image Analysis Application
- 3. Statistical Approach: Background
- 4. Statistical Approach: Preliminary Results
- 5. Current Directions

Foci Analysis Application



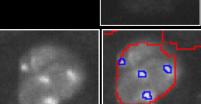


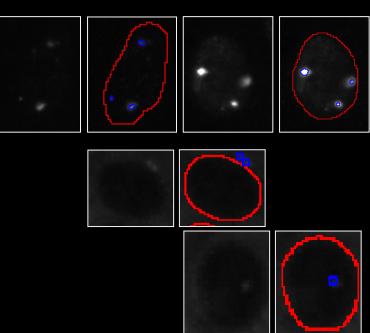
Foci Analysis Application



Pros

- Achieves high-accuracy foci detection for variety of large image sets
- Automatic collection of foci statistics (size, intensity, contrast, quantity per nucleus)
- Currently in use by radiation biologists (Costes, et. al., Bioastronautics Investigators' Workshop, 2005)



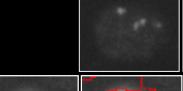


Foci Analysis Application

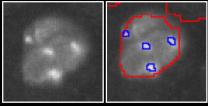


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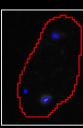
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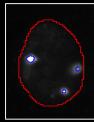






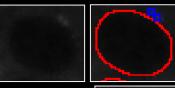




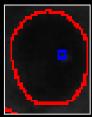


Cons

- Parameters adjusted by user can bias results
- Designed for a specific type of feature
- Toolbox approach not easily extendable to other tasks

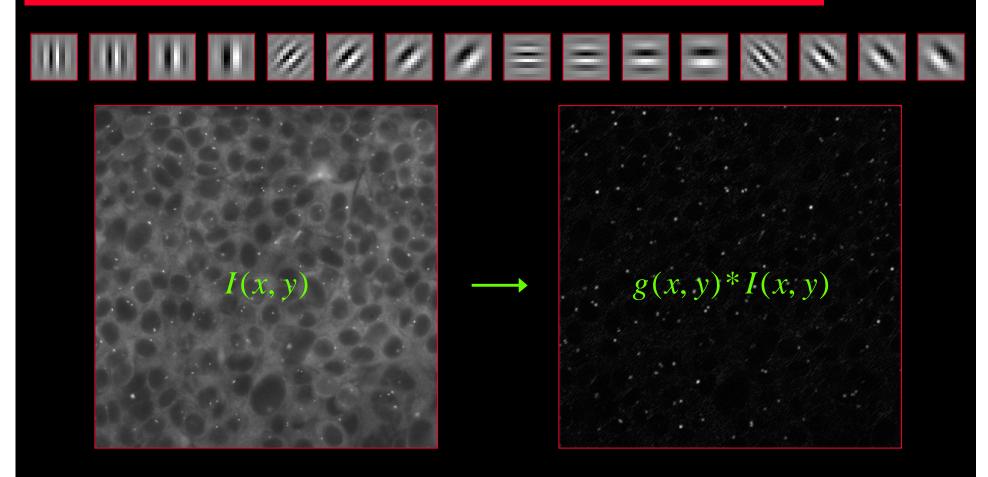






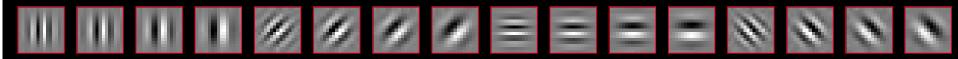
Feature Detection via Filter Banks

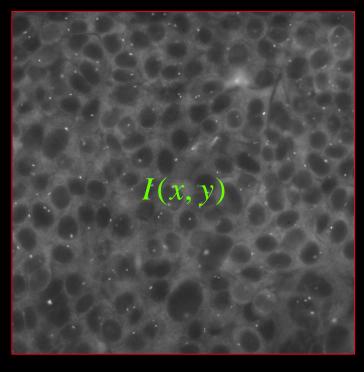




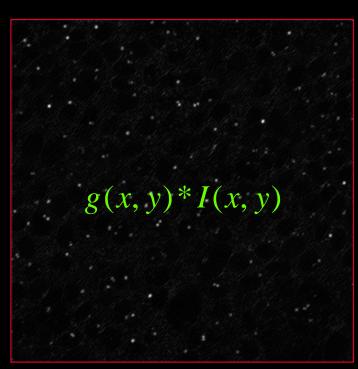
Feature Detection via Filter Banks







Gabor wavelet transform



$$g(x, y, \theta, \lambda) =$$

$$\exp\left[-\frac{1}{2}\left\{\frac{(x\cos\theta+y\sin\theta)^2}{\sigma_x^2}+\frac{(-x\sin\theta+y\cos\theta)^2}{\sigma_y^2}\right\}\right]\exp\left[\frac{2\pi(x\cos\theta+y\sin\theta)}{\lambda}i\right]$$

March 5, 2005

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How to Learn Basis From Data?



Independent Components Analysis (Comon, 1991)

- Use statistical criteria to define optimal basis for decomposing images into combinations of basis functions
- Optimization yields multi-oriented, multiscale, bandpass basis functions specifically tuned to data set

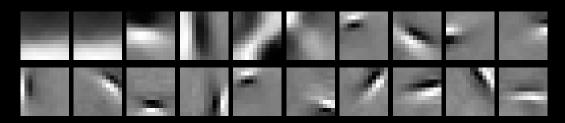
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example basis functions



Sparse Coding (Olshausen-Field, 1996) Independents Components Analysis (Bell & Sejnowski, 1997)

 More localized than Fourier filters, arbitrary frequencies and orientations, unlike Gabor and wavelet filters

Independent Components Analysis



 Consider an arbitrary NxN subimage of a sample image for fixed N:

Independent Components Analysis



- Consider an arbitrary NxN subimage of a sample image for fixed N:
- Treat each subimage as an observed vector variable
 x that is a mixture of unknown basis functions a_i with
 unknown coefficients s_i

$$\mathbf{x} = s_1 \mathbf{a}_1 + s_2 \mathbf{a}_2 + \dots + s_n \mathbf{a}_n$$

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- Unknowns: basis functions a and coefficients s;
- Given: only observations x

$$x = As$$

Optimization



$$\mathbf{x} = \mathbf{A}\mathbf{s}$$

Express unknown coefficients in terms of inverse W of unknown mixing matrix A

$$\mathbf{s}^* = \mathbf{W}\mathbf{x}$$

Coefficient estimates §* are called independent components

Optimization



$$\mathbf{x} = \mathbf{A}\mathbf{s}$$

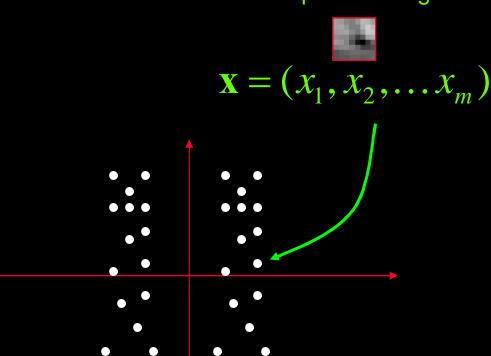
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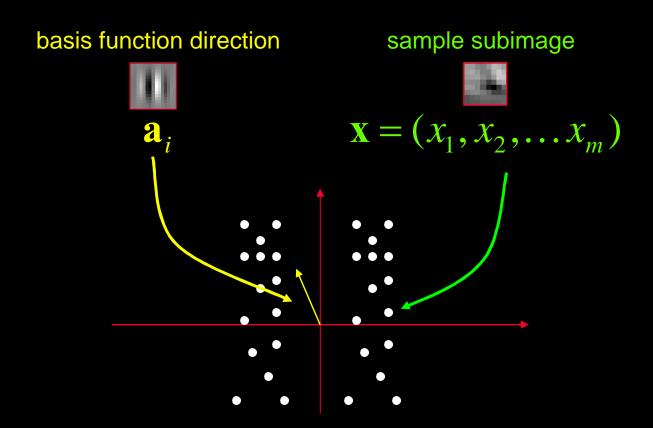
- Coefficient estimates §* are called independent components
- Optimize a function J(s*) that encourages the independent components to be as non-Gaussian as possible



sample subimage

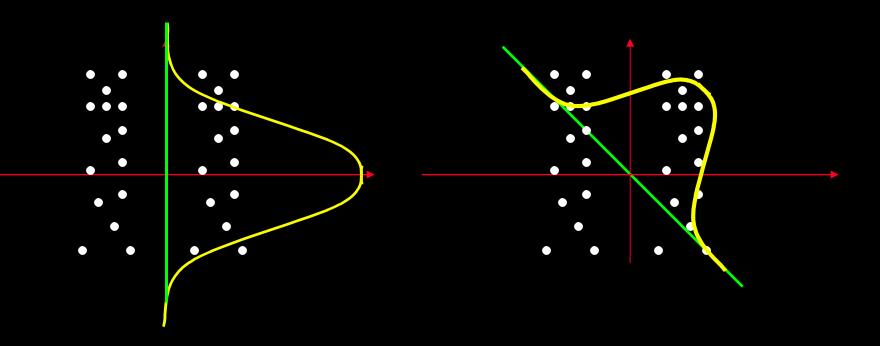






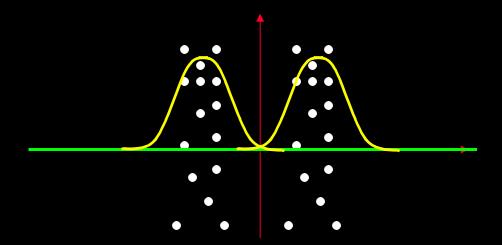


 Projection pursuit: random subspace projections of structured point clouds are typically Gaussian



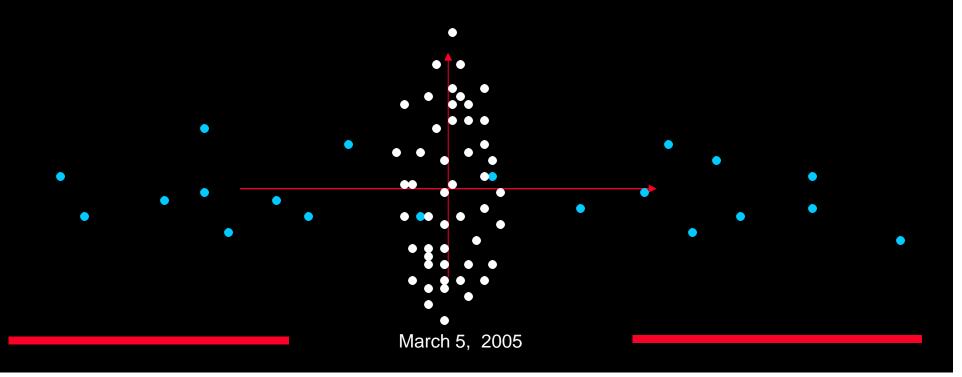


- Projection pursuit: random subspace projections of structured point clouds are typically Gaussian
- Interesting projections are not Gaussian, e.g., mixture of Gaussians for cluster discrimination

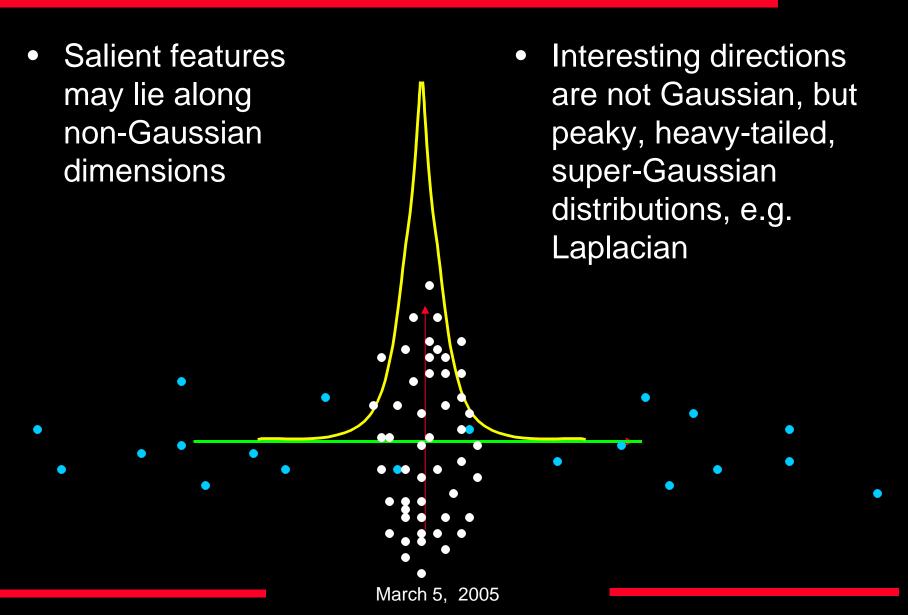




 Salient features may lie along non-Gaussian dimensions







Recall: Optimization Constraints



$$\mathbf{x} = \mathbf{A}\mathbf{s}$$

- Optimize a function $J(s^*)$ that encourages the independent components to be as *non-Gaussian* as possible
- Output: basis functions and independent components

$$\mathbf{x} = s_1 \mathbf{a}_1 + s_2 \mathbf{a}_2 + \cdots + s_n \mathbf{a}_n$$

Non-Gaussianity: Contrast Functions



Entropy: measure of how random vs. structured a random variable is

$$H(\mathbf{y}) = -\int f(\mathbf{y}) \log f(\mathbf{y}) d\mathbf{y}$$

Negentropy: difference in entropy w.r.t. Gaussian with same variance

$$J(\mathbf{y}) = H(\mathbf{y}_{\text{gauss}}) - H(\mathbf{y})$$

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Negentropy Approximation:

- c constant
- v standard Gaussian variable
- G non-quadratic function

$$J(\mathbf{y}) \approx$$

$$c[E\{G(\mathbf{y})\} - E\{G(\mathbf{v})\}]^2$$

FastICA: A. Hyvärinen. Fast and Robust Fixed-Point Algorithms for Independent Component Analysis. *IEEE Transactions on Neural Networks* 1999.

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Independent Components of Microscopy Imagery



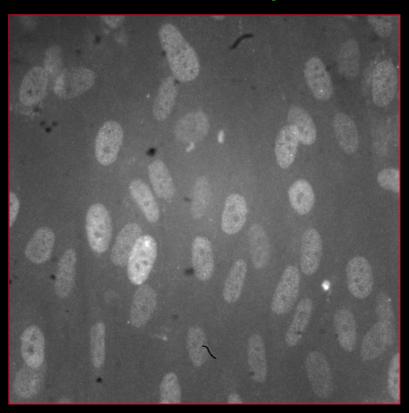
Goals

- Represent salient features characterizing cell differences, e.g. foci
- Distinguish differently treated cells and images
- Design task-specific algorithms: feature extraction, cell and image classification

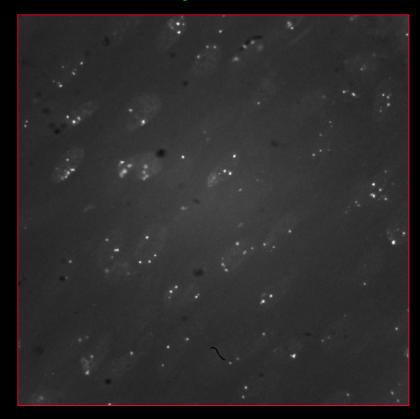
Preliminary Test



Control Group



Dose: 30cGy, Time: 10min

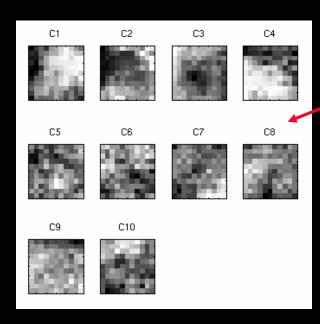


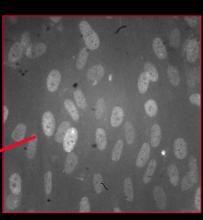
- Input data: 15x15 pixel subimages within nuclei
- Centered at local maxima
- Small training set (~1000 subimages each)

Example Microscopy Basis Functions



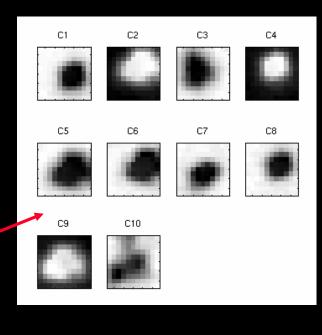
Basis Functions Control Group







Basis Functions Dose: 30cGy, Time: 10min



- Subimages centered and whitened (decorrelated, unit variance)
- Data dimensionality may be reduced by PCA
- Basis functions defined up to unknown sign ambiguity

Large Data Set

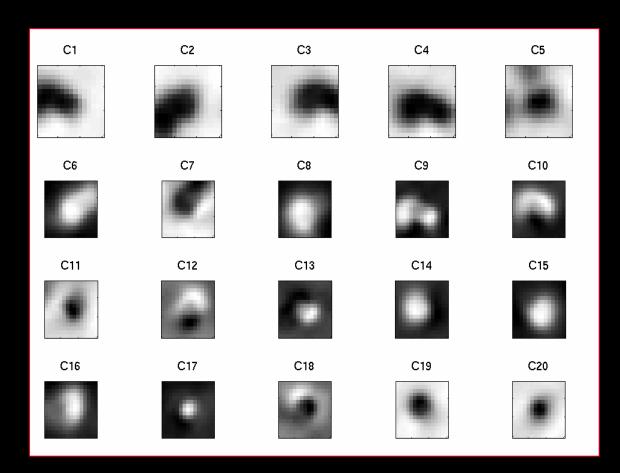


- ~100 1.3 Mb images
- 9 classes (varying dose/time treatment groups)
- ~10 images per class2 training, 8 testing
- ~10,000 15x15 subimages
- random sample of ~2000 training examples
- dimensionality reduction by PCA from 225 to 20

Large Data Set



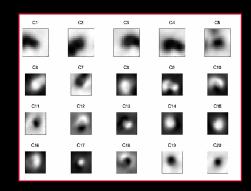
Basis Functions

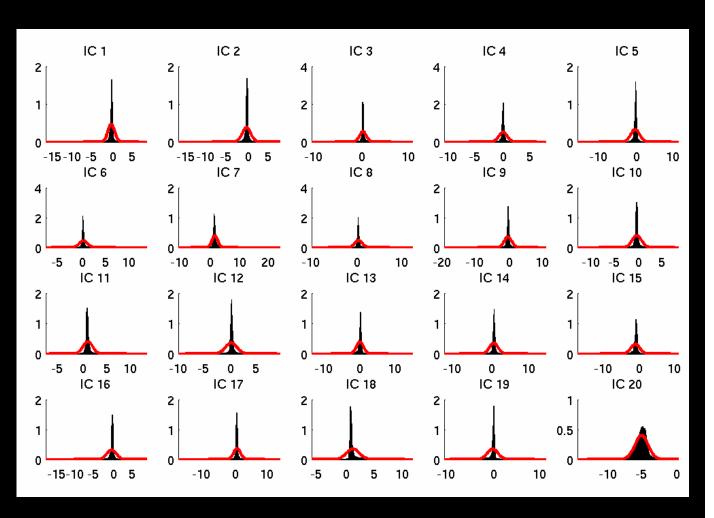


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Independent Component Distributions



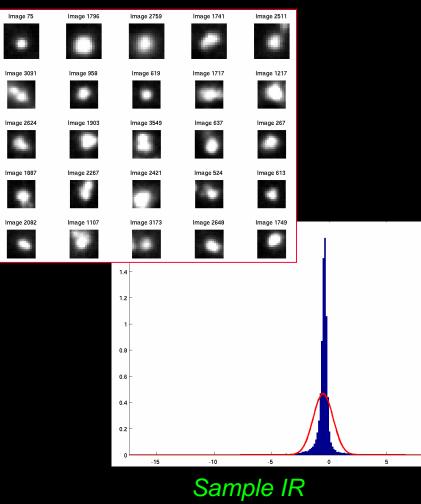




Classification by Maximum Response

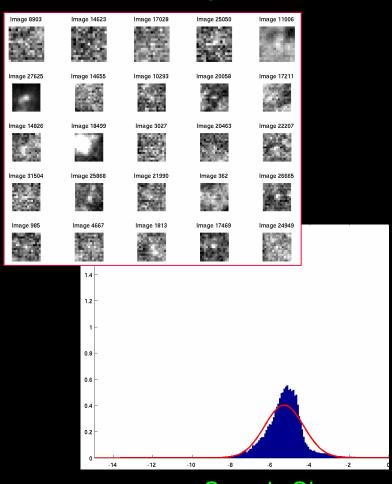


Sample Subimages: FG Class



Nucleus

Sample Subimages: BG Class



Sample Sham **Nucleus**

Classification by Maximum Response

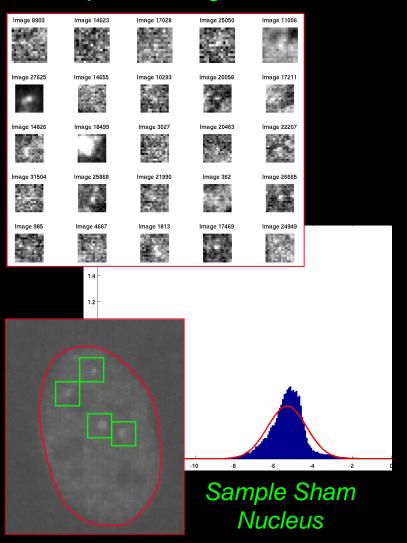


Sample Subimages: FG Class

Sample IR

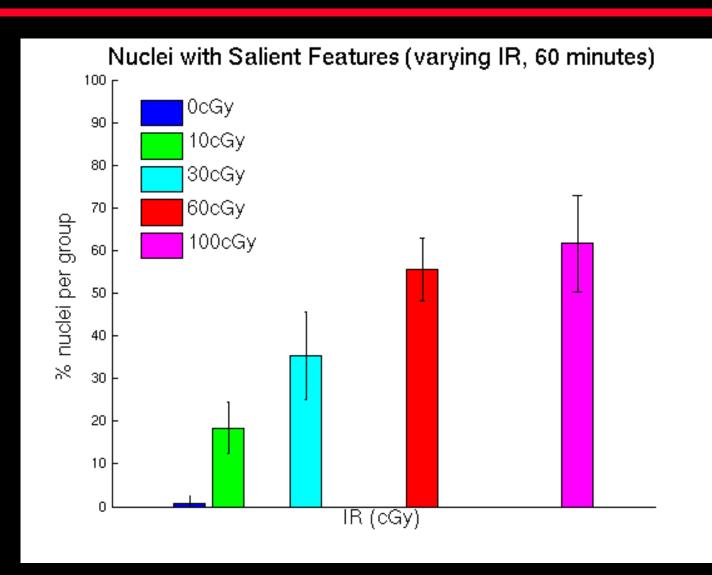
Nucleus

Sample Subimages: BG Class



Validation





Conclusions and Current Directions



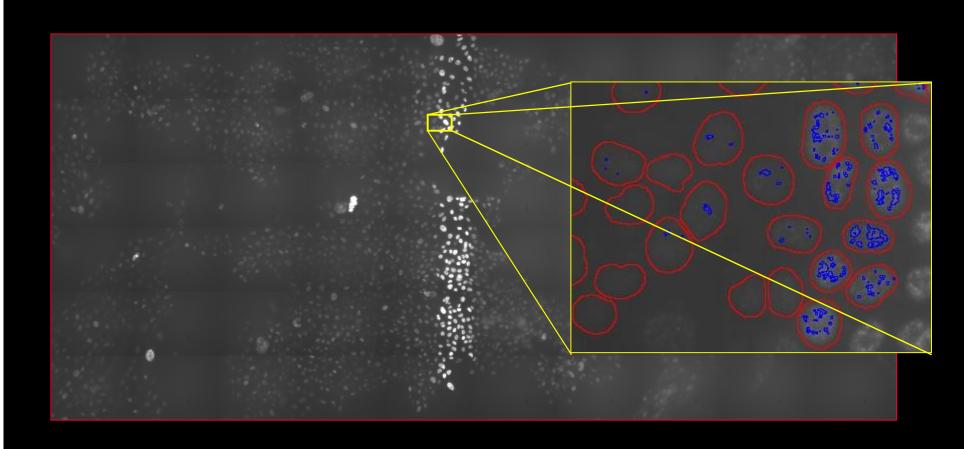
- Promising statistical method for learning representation of salient features.
- Away from hand-designed, problem-specific tools, toward general purpose solutions.

Next

- Applications to broad variety of imagery
- Feature extraction algorithms
 - subimage classification for feature detection
 - multiscale analysis
 - validation of accuracy/precision
- Cell and image classification: Bayesian modeling of treatment groups

Future Work: Bystander Effect







Thank You

Current Directions: Engineering



- Usability: trade-off between flexibility and simplicity.
 e.g. trainable vs. interactive vs. fully automatic
- Cross-platform implementation integrating analysis, visualization, browsing
- VTK/ITK open source commercial-grade libraries developed in medical imaging community
- Capability for distributed and parallel computing for large data sets

Feature Detection via Filter Banks



$$g(x, y, \theta, \lambda) =$$

$$\exp\left[-\frac{1}{2} \left\{ \frac{(x\cos\theta + y\sin\theta)^2}{\sigma_x^2} + \frac{(-x\sin\theta + y\cos\theta)^2}{\sigma_y^2} \right\} \right]$$

$$\exp\left[\frac{2\pi(x\cos\theta + y\sin\theta)}{\lambda}i\right]$$

Parameters:

 σ_x : Gaussian standard deviation in x-direction

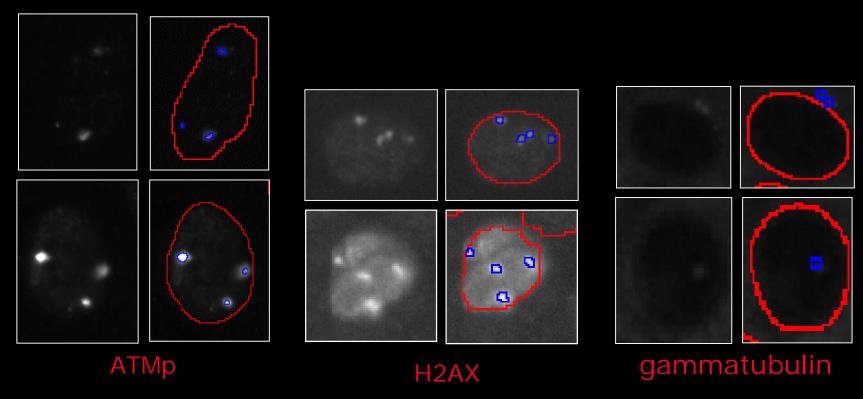
σ_y: Gaussian standard deviation in y-direction

∴ sinusoid wavelength

: filter orientation

Image Processing Approach

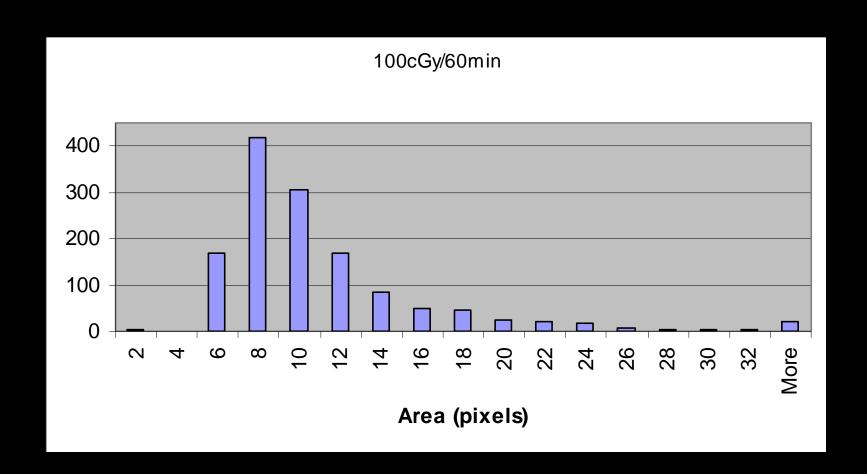




- Geometric nucleus segmentation provides local context
- Foci detection via intensity statistics and locally adaptive thresholds
- User preferences for adapting software to data set

Scientific Results: Size and Number of Foci

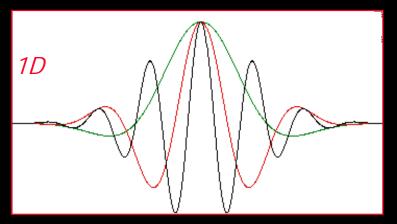


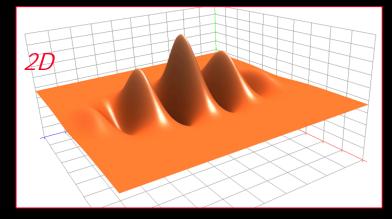


Feature Detection via Filter Banks



Gabor Wavelet Function sinusoid modulated by Gaussian envelope





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FastICA Implementation



- A. Hyvärinen. Fast and Robust Fixed-Point Algorithms for Independent Component Analysis. *IEEE Transactions on Neural Networks* 10(3):626-634, 1999.
- Maximizes approximation to negentropy: easy to compute, compromise between negentropy and kurtosis
- Data pre-whitened and reduced with classical PCA
- Fixed-point optimization, block-mode computation
- Parallel and distributed
- Cubic convergence rate
- c is a constant, v is a zero-mean, unit-variance Gaussian variable, G is a non-quadratic function

$$J(\mathbf{y}) \approx c[E\{G(\mathbf{y})\} - E\{G(\mathbf{v})\}]^2$$

Features with High Response



Control Group

Image 358 Image 432 Image 545 Image 552 Image 553

Image 560 Image 561 Image 562 Image 563 Image 565

Image 566 Image 587 Image 594 Image 595 Image 601

Image 702

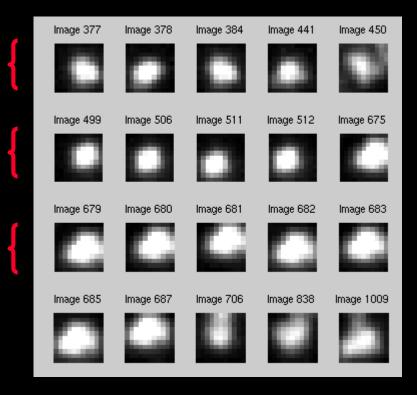
Image 730

Image 975

Image 606

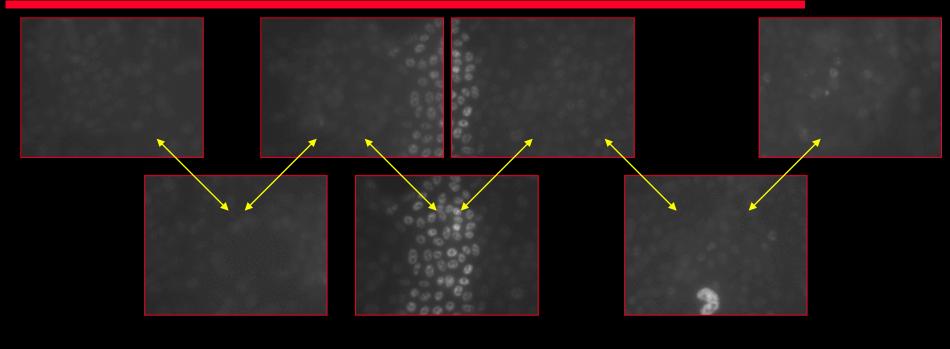
Image 659

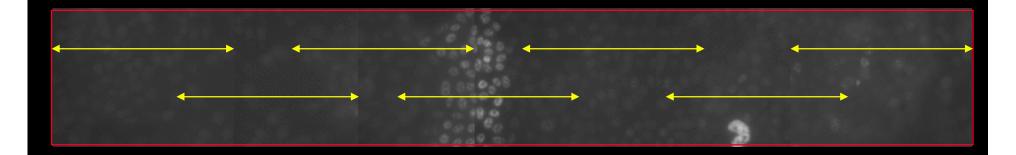
Dose: 30cGy Time: 10 minutes



Bystander Experiment

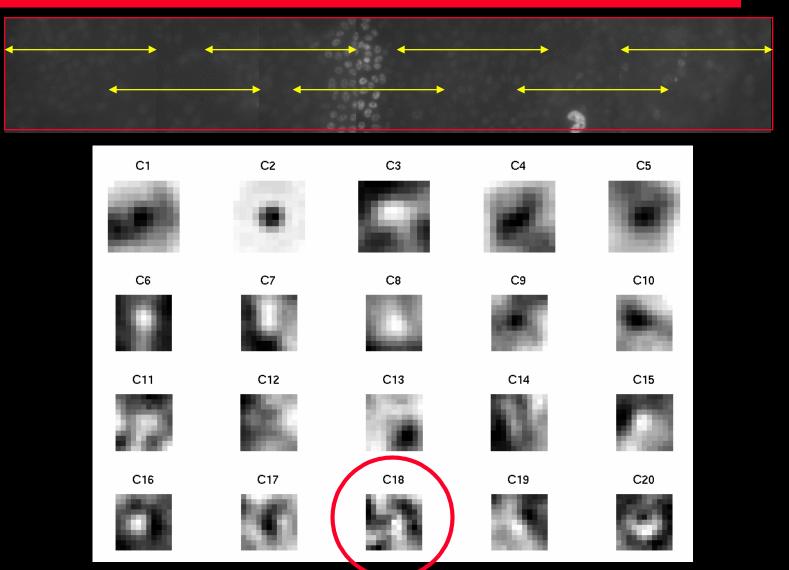






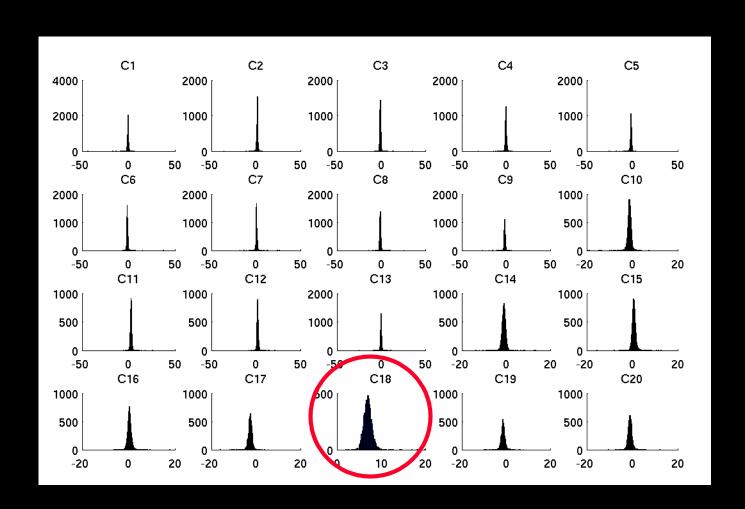
Bystander Experiment





Response Histograms



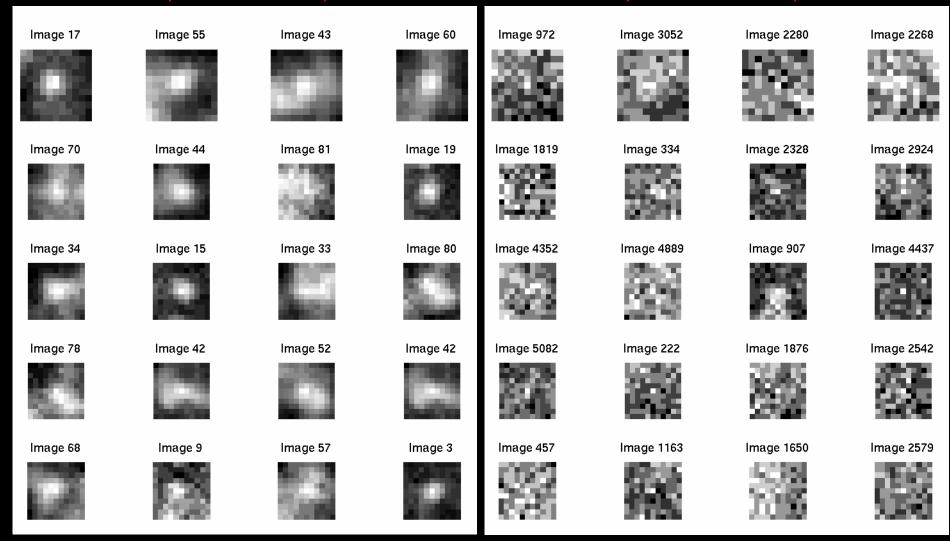


Classification by Maximum Response



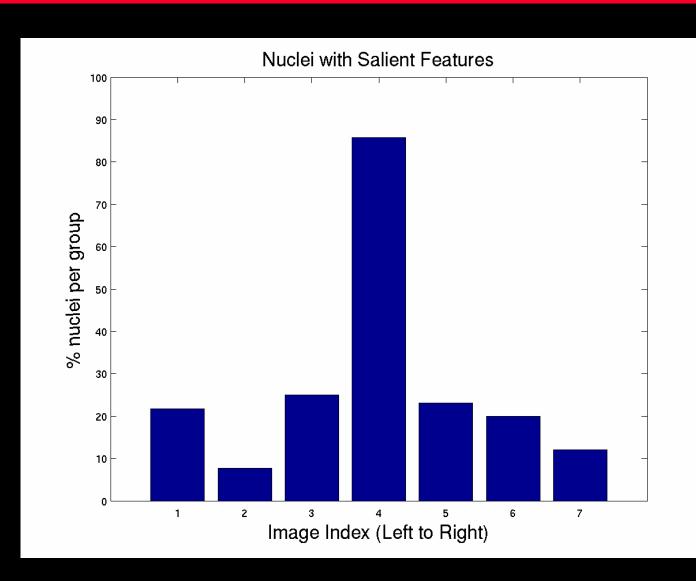
Sample Images: Class 1 (less Gaussian)

Sample Images: Class 2 (more Gaussian)



Validation



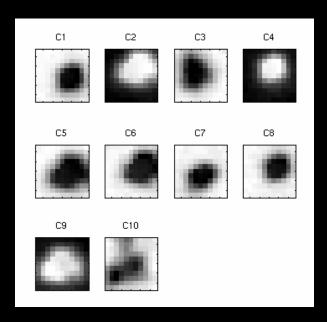


Learned Basis vs. Filter Bank

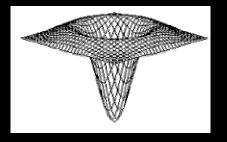


How do learned basis functions compare to typical filter bank kernels?

Learned Basis Functions



2D Laplacian of Gaussian



2D Gabor Kernel

